

Cytokine Levels in Critically Ill Patients With COVID-19 and Other Conditions

An abnormally strong proinflammatory response known as “cytokine storm” may play an important role in the pathophysiology of coronavirus disease 2019 (COVID-19), although cytokine storm remains ill defined.¹ Sinha and colleagues² reported that although IL-6 levels are elevated in severe COVID-19, they are lower than levels usually observed in (non-COVID-19) acute respiratory distress syndrome (ARDS). However, this comparison is limited by the use of different assays, which are not well standardized.³ We compared cytokine levels in critically ill patients with COVID-19 vs levels in patients with other critical illnesses.

Methods | All patients in this study were admitted to the intensive care unit (ICU) of Radboud University Medical Center. Plasma concentrations of the proinflammatory cytokines tumor necrosis factor (TNF), IL-6, and IL-8 were determined in consecutive mechanically ventilated patients with COVID-19 with ARDS (partial pressure of oxygen/fraction of inspired oxygen ratio <300; sampled within 48 hours after ICU admission), bacterial septic shock with or without ARDS (sampled within 24 hours after septic shock diagnosis), out-of-hospital cardiac arrest (OHCA; sampled within 24 hours after ICU admission), and multiple traumas (sampled within 24 hours after trauma). The patients with sepsis and trauma are part of larger published cohorts,^{4,5} whereas data of 14 patients with OHCA were previously published.⁶ Sampling

occurred between 2010 and 2020 (Table). Patients with immunological insufficiencies were excluded, defined as chronic/concomitant use of immunosuppressive medication, chemotherapy/radiotherapy in the last year or in the past for (non-)Hodgkin lymphoma, or humoral/cellular deficiencies. Cytokines in all cohorts were determined using the same methodology (Milliplex assay, Millipore, on a MAGPIX instrument, Luminex Corporation) by the same technician using the same protocol.

Patient characteristics were analyzed using Fisher exact or Kruskal-Wallis tests followed by Dunn post hoc tests. Cytokine data are presented as geometric means (95% CIs) and analyzed using 1-way analysis of variance on log-transformed data followed by Dunnett post hoc tests. Data were analyzed using Graphpad Prism version 8.3.0 (Graphpad Software). A 2-sided $P < .05$ was considered statistically significant. The study was carried out in accordance with the applicable rules concerning the review of research ethics committees and informed consent in the Netherlands. All patients or legal representatives were informed about the study details and allowed to abstain from participation. Patients who consented to participate or their next of kin provided oral consent.

Results | There were 46 patients with COVID-19 with ARDS, 51 with septic shock with ARDS, 15 with septic shock without ARDS, 30 with OHCA, and 62 with multiple traumas. There were no significant differences in sex or age between patients with COVID-19 and other patient groups (Table). Patients with

Table. Patient Characteristics^a

Characteristic	COVID-19 with ARDS, March 11 to April 27, 2020 (n = 46)	Septic shock, March 15, 2013, to March 28, 2017		Out-of-hospital cardiac arrest, February 5, 2010, to December 12, 2013 (n = 30)	Trauma, March 19, 2011, to May 30, 2013 (n = 62)
		With ARDS (n = 51)	Without ARDS (n = 15)		
Sex, No. (%)					
Male	34 (74)	36 (71)	6 (40)	22 (73)	44 (71)
Female	12 (26)	15 (29)	9 (60)	8 (27)	18 (29)
Age, median (IQR), y	67 (57-71)	62 (53-72)	73 (64-78)	65 (52-75)	58 (37-72)
BMI, median (IQR)	27.5 (25.0-29.3)	26.4 (23.8-30.5)	25.0 (21.5-30.3)	25.1 (23.4-26.9) ^b	24.7 (23.2-27.4) ^c
Medical history, No. (%)					
Cardiovascular insufficiency	12 (26)	2 (4) ^c	2 (13)	1 (3) ^b	1 (2) ^d
Respiratory insufficiency	3 (7)	1 (2)	0	0	0
COPD	3 (7)	5 (10)	0	0	0
Kidney insufficiency	0	5 (10)	0	0	0
Metastatic neoplasm	4 (9)	1 (2)	2 (13)	1 (3)	0 ^b
Diabetes	13 (28)	8 (16)	1 (7)	1 (3) ^c	4 (6) ^c
Hematologic malignancy	0	0	0	0	0
APACHE II score, median (IQR) ^e	14 (12-18)	21 (17-26) ^d	24 (18-31) ^d	27 (20-34) ^d	20 (14-25) ^c
Pao ₂ /Fio ₂ ratio, median (IQR)	139 (107-171)	206 (162-260) ^d	354 (328-424) ^d	246 (159-370) ^d	253 (201-361) ^d
Leukocytes, median (IQR), ×10 ⁹ /L	8.2 (6.4-11.1)	14.0 (9.8-20.8) ^d	15.4 (7.2-24.4) ^c	12.9 (10.0-16.7) ^d	11.8 (8.9-14.0) ^c

Abbreviations: APACHE II, Acute Physiology and Chronic Health Evaluation II; ARDS, acute respiratory distress syndrome; BMI, body mass index (calculated as weight in kilograms divided by height in meters squared); COPD, chronic obstructive pulmonary disease; COVID-19, coronavirus disease 2019; Fio₂, fraction of inspired oxygen; IQR, interquartile range; Pao₂, partial pressure of oxygen.

^a Data were obtained on the same day that blood was obtained for cytokine determination.

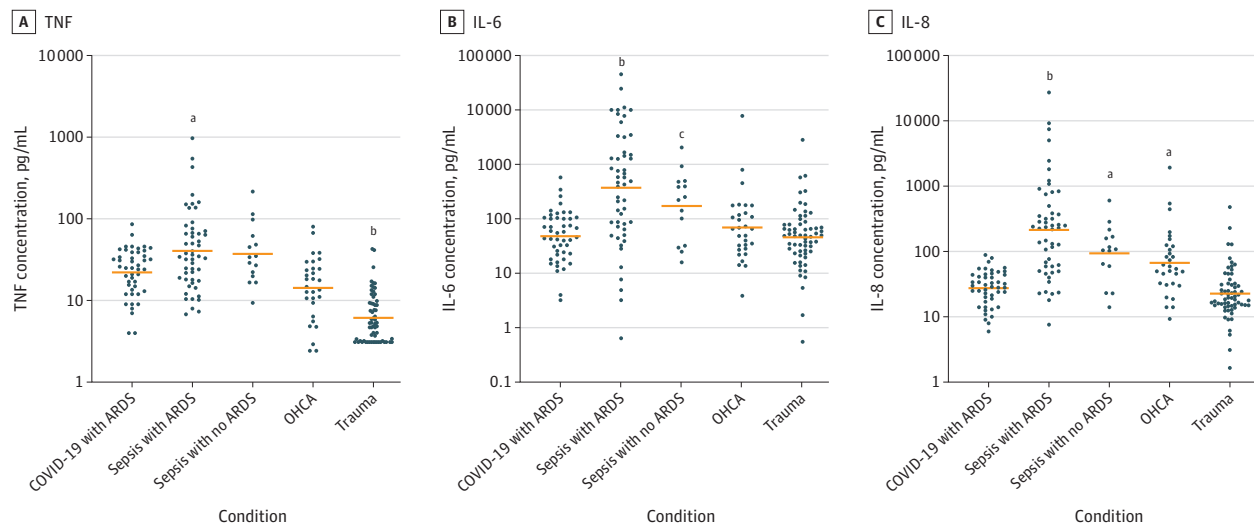
^b $P < .05$ vs COVID-19 with ARDS.

^c $P < .01$ vs COVID-19 with ARDS.

^d $P < .001$ vs COVID-19 with ARDS.

^e Intensive care unit score of overall disease severity ranging from 0 to 71; a higher score indicates more severe disease.

Figure. Cytokine Levels in Critically Ill Patients With Coronavirus Disease 2019 (COVID-19) and Other Conditions



Plasma concentrations of tumor necrosis factor (TNF) (A), IL-6 (B), and IL-8 (C) in patients with COVID-19 and acute respiratory distress syndrome (ARDS) (n = 46), septic shock with ARDS (n = 51), septic shock without ARDS (n = 15), out-of-hospital cardiac arrest (OHCA; n = 30), and multiple traumas (n = 62).

Data are presented as scatter plots with red horizontal bars indicating the geometric mean levels.

^a $P < .01$ vs COVID-19 with ARDS.

^b $P < .001$ vs COVID-19 with ARDS.

^c $P < .05$ vs COVID-19 with ARDS.

COVID-19 had a higher body mass index and prevalence of diabetes than patients with OHCA and trauma. In COVID-19, cardiovascular insufficiency was more common, overall disease severity and leukocyte counts were lower, and lung injury was more severe compared with the other groups.

Levels of all 3 cytokines were significantly lower in patients with COVID-19 than in patients with septic shock with ARDS; the geometric means were 22 pg/mL (95% CI, 18-27) vs 40 pg/mL (95% CI, 30-55) ($P < .01$) for TNF; 48 pg/mL (95% CI, 35-66) vs 376 pg/mL (95% CI, 190-744) ($P < .001$) for IL-6; and 27 pg/mL (95% CI, 23-33) vs 215 pg/mL (95% CI, 133-347) ($P < .001$) for IL-8 (depicted in the Figure on a log scale). Patients with COVID-19 also displayed significantly lower IL-6 and IL-8 concentrations compared with patients with septic shock without ARDS (Figure). TNF levels in patients with COVID-19 were higher than those in trauma patients, whereas no differences between patients with COVID-19 and OHCA or trauma were present for IL-6. For IL-8, lower concentrations were found in patients with COVID-19 compared with patients with OHCA, while no differences vs the trauma group were observed.

Discussion | In this study, critically ill patients with COVID-19 with ARDS had circulating cytokine levels that were lower compared with patients with bacterial sepsis and similar to other critically ill patients. These findings are in line with lower leukocyte counts observed in patients with COVID-19, and are possibly due to lower overall disease severity, despite the presence of severe pulmonary injury. The findings of this preliminary analysis suggest COVID-19 may not be characterized by cytokine storm. Whether anticytokine therapies will benefit patients with COVID-19 remains to be determined. Limi-

tations of the study include the small sample sizes, single center involved, and the use of different lots of the same assays without data on lot-to-lot variability.

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Concept and design: Kox, Pickkers.

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1. Tisoncik JR, Korth MJ, Simmons CP, Farrar J, Martin TR, Katze MG. Into the eye of the cytokine storm. *Microbiol Mol Biol Rev.* 2012;76(1):16-32. doi:10.1128/MMBR.05015-11

2. Sinha P, Matthay MA, Calfee CS. Is a "cytokine storm" relevant to COVID-19? *JAMA Intern Med.* Published online June 30, 2020. doi:10.1001/jamainternmed.2020.3313

3. Khan SS, Smith MS, Reda D, Suffredini AF, McCoy JP Jr. Multiplex bead array assays for detection of soluble cytokines: comparisons of sensitivity and quantitative values among kits from multiple manufacturers. *Cytometry B Clin Cytom*. 2004;61(1):35-39. doi:10.1002/cyto.b.20021
4. Habes QLM, van Ede L, Gerretsen J, Kox M, Pickkers P. Norepinephrine contributes to enterocyte damage in septic shock patients: a prospective cohort study. *Shock*. 2018;49(2):137-143. doi:10.1097/SHK.0000000000000955
5. Timmermans K, Kox M, Vaneker M, et al. Plasma levels of danger-associated molecular patterns are associated with immune suppression in trauma patients. *Intensive Care Med*. 2016;42(4):551-561. doi:10.1007/s00134-015-4205-3
6. Timmermans K, Kox M, Gerretsen J, et al. The involvement of danger-associated molecular patterns in the development of immunoparalysis in cardiac arrest patients. *Crit Care Med*. 2015;43(11):2332-2338. doi:10.1097/CCM.0000000000001204

Racial/Ethnic Variation in Nasal Gene Expression of Transmembrane Serine Protease 2 (TMPRSS2)

Coronavirus disease 2019 (COVID-19) has disproportionately affected communities of color.^{1,2} In many areas of the US, infection and death rates for COVID-19 are 2 to 3 times higher in Black individuals than their proportion of the population.^{1,2} Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is spread by airway contact and uses transmembrane serine protease 2 (TMPRSS2) to facilitate viral entry and spread.³ Host-expressed TMPRSS2 on nasal and bronchial epithelium activates the SARS-CoV-2 spike protein and cleaves the angiotensin-converting enzyme 2 receptor to which the virus binds, enabling SARS-CoV-2 to enter the body.³

Racial/ethnic differences in TMPRSS2 gene-related activity in prostate tissue have been associated with disproportionately higher incidence of prostate cancer in Black men vs White men.⁴ Recognizing that many factors contribute to COVID-19 health disparities, we investigated TMPRSS2 nasal gene expression in a racially/ethnically diverse cohort.

Methods | This cross-sectional study used nasal epithelium collected during 2015-2018 from individuals within the Mount Sinai Health System (New York, New York), a cohort we have previously studied.⁵ Healthy individuals and individuals with asthma aged 4 to 60 years underwent nasal brushing for research on asthma biomarkers.

Self-identified race/ethnicity was queried given prior associations between race/ethnicity and asthma. RNA isolation of brushings followed by RNA sequencing, sequence alignment, and normalization were performed. The Mount Sinai institutional review board approved the study. Written informed consent was obtained from participants.

Linear regression modeling adjusted for age, sex, and asthma with TMPRSS2 expression in log₂ counts per million as the dependent variable and self-identified race/ethnicity as

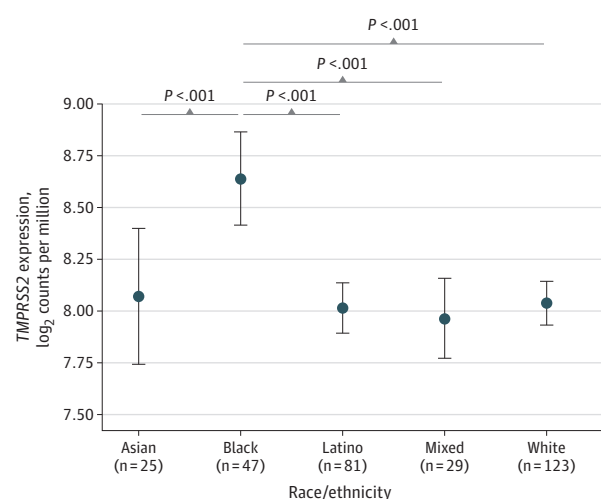
the independent variable was performed using R version 3.6.0 (R Foundation for Statistical Computing). Two-sided tests and a significance threshold of $P \leq .05$ were used.

Results | The cohort (n = 305) was 8.2% Asian individuals, 15.4% Black individuals, 26.6% Latino individuals, 9.5% individuals of mixed race/ethnicity, and 40.3% White individuals. Of the participants, 48.9% were male and 49.8% had asthma.

Among the racial/ethnic groups, nasal gene expression of TMPRSS2 was highest in Black individuals (n = 47; mean, 8.64 [95% CI, 8.41-8.86] log₂ counts per million) compared with Asian individuals (n = 25; mean, 8.07 [95% CI, 7.74-8.40] log₂ counts per million), Latino individuals (n = 81; mean, 8.02 [95% CI, 7.90-8.14] log₂ counts per million), individuals of mixed race/ethnicity (n = 29; mean, 7.97 [95% CI, 7.77-8.16] log₂ counts per million), and White individuals (n = 123; mean, 8.04 [95% CI, 7.94-8.15] log₂ counts per million) (Figure).

TMPRSS2 expression was significantly higher in Black individuals compared with Asian, Latino, mixed race/ethnicity, and White individuals (all $P < .001$) based on linear regression (Figure and Table). There were no significant associations between TMPRSS2 expression and sex, age, or asthma.

Figure. Nasal Gene Expression of TMPRSS2 in Self-identified Racial/Ethnic Groups



The data points indicate means and the error bars indicate 95% CIs for transmembrane serine protease 2 (TMPRSS2) gene expression in self-identified racial/ethnic groups. The P values were calculated using linear regression modeling in which TMPRSS2 gene expression was the dependent variable and race/ethnicity was the independent variable.

Table. β Coefficients for Race/Ethnicity From Linear Regression Modeling^a

Race/ethnicity	Unadjusted β coefficient (95% CI) ^b	P value	Adjusted β coefficient (95% CI) ^{b,c}	P value
Black	[Reference]		[Reference]	
Asian	-0.57 (-0.87 to -0.27)	<.001	-0.63 (-0.94 to -0.32)	<.001
Latino	-0.62 (-0.85 to -0.40)	<.001	-0.64 (-0.86 to -0.42)	<.001
Mixed race	-0.67 (-0.96 to -0.39)	<.001	-0.66 (-0.95 to -0.37)	<.001
White	-0.60 (-0.81 to -0.39)	<.001	-0.60 (-0.81 to -0.39)	<.001

^a TMPRSS2 expression was the dependent variable and self-identified race/ethnicity was the independent variable.

^b β coefficients indicate the difference in TMPRSS2 expression in log₂ counts per million between a given race/ethnicity and Black individuals.

^c Adjusted for age, sex, and asthma.